

## SPECIFICATION

INSAI

A Membrane Protein Polypeptide Having Pre-B Cell Growth-Supporting Ability and A Gene Thereof

### Technical Field

The present invention relates to a gene and a novel membrane protein encoded by said gene, and more specifically, relates to a gene encoding a novel membrane protein polypeptide having pre-B cell growth-supporting ability, a vector containing said gene, transformants transformed by said vector and a method for producing the novel membrane protein polypeptide by using said gene.

The present invention further relates to a monoclonal antibody recognizing a novel membrane protein polypeptide having pre-B cell growth-supporting ability.

The gene of the present invention encodes a novel membrane protein polypeptide enhancing pre-B cell growth-supporting ability on the surface of synovial cells derived from patients with rheumatoid arthritis (RA). In the present invention, a homogeneous and purified novel membrane protein polypeptide having pre-B cell growth-supporting ability can be produced in large quantities by transforming appropriate host cells with a suitable vector in which the gene of the present invention is inserted. Thus, according to the present invention, it becomes possible to identify rheumatoid arthritis (RA), and also prepare reagents for the clinical diagnosis thereof.

### Background Art

Inflammatory cells in the synovial membrane and the synovial fluid of patients with rheumatoid arthritis (RA) are derived from peripheral blood and the migration

of these cells to synovial membranes has not been explicated perfectly yet, but it is believed to be caused by a complicated interaction between chemical signals given to cells and protein (adhesion molecule) on cell membranes.

Various studies upon the significance of membrane proteins in arthritis have been performed. For example, it is known that an intercellular adhesion molecule-1 (hereinafter referred to as ICAM-1) is expressed on the inner layer of the synovial membrane and the blood vessel of the synovial membrane of patients with rheumatoid arthritis (RA), which is a ligand of a T cell surface molecule LFA-1 and causes both adhesion and migration of cells in the blood vessel wall [Hale et al.; Arthritis Rheum., 32:22 (1989), and Haynes et al.; Springer Semin. Immunopathol., 11:163 (1989) ] .

Similarly, it is suggested that a vasocellular adhesion molecule-1 (hereinafter referred to as VCAM-1), which is a ligand of integrin VLA-4 expressed on T lymphoid cells (memory cells in particular) and monocytes, is expressed on the synovial membrane and fibroblast-like synovial cells of rheumatoid arthritis (RA) and osteoarthritis, [Morales-Ducret et al.; J. Immunol., 149:1424 (1992) ] , and further that a membrane protein called VAP-1 is expressed on the endothelial vein of a synovial membrane and may work as a specific recognition structure of leukocytes [Salmi et al.; Science, 257, 1407 (1992)] .

The present inventors have engaged in extensive studies with a view to investigating the function of the bone marrow microenvironments in disorders causing abnormalities of B cells, and have found that the pre-B cell growth-supporting ability of bone marrow stromal cells derived from patients with rheumatoid arthritis (RA) and multiple myeloma (MM) is enhanced in

comparison with that of healthy donor-derived bone marrow stromal cells and that the direct contact of pre-B cells with stromal cells might play an essential role in this supporting ability. And the present inventors have established novel stromal cell lines (RASV5-5, MMSV3-3) containing a molecule enhancing the growth of pre-B cells by cell-lining stromal cells of patients, and have found that the pre-B cell growth-supporting activity of these stromal cell lines is most likely caused by unknown adhesion molecules different from known stem cell factors (SCF), ICAM, CD44, VCAM-1, LFA-1 $\alpha$ , LFA-1 $\beta$ , NCAM and FLAM-1 [J. Immunol., 149:4088 (1992)] .

Further, since it has been suggested that the synovial cell line SynSV6-14 established from the synovial cell derived from patients with rheumatoid arthritis (RA) has pre-B cell growth-supporting ability similarly to the stromal cell line RASV5-5 derived from the bone marrow of patients with rheumatoid arthritis (RA), the present inventors have obtained a novel monoclonal antibody which responds to these cell lines but does not respond to the stromal cell line NFSV1-1 derived from the human bone marrow having no pre-B cell growth-supporting ability, and at the same time have succeeded in cloning genes encoding its antigen membrane protein (Bst-1) (Japanese Patent Application No. 5-141178/1993).

#### Disclosure of Invention

The present inventors have obtained a novel mouse monoclonal antibody RS38 which responds to SynSV6-14 but does not respond to the healthy bone marrow stromal cell line NFSV1-1 and recognizes a membrane protein different from the above Bst-1 at the process of producing various mouse monoclonal antibodies recognizing a membrane protein expressed on the

synovial cell derived from patients with rheumatoid arthritis (RA) but not expressed on the cell derived from healthy donors. Subsequently, the present inventors have succeeded in isolating clones encoding a novel membrane protein responding to said RS38, according to screening a cDNA library prepared from a synovial cell line derived from patients with rheumatoid arthritis (RA) by using the RS38 antibody, which has led to the completion of the present invention.

That is, the present invention is directed to provide a novel membrane protein polypeptide having pre-B cell growth-supporting ability, a gene encoding said polypeptide, a vector containing said gene, transformants transformed by said vector and a method for producing a novel membrane protein by using said gene.

Further, the present invention is directed to provide a monoclonal antibody recognizing a novel membrane protein having pre-B cell growth-supporting ability.

The present invention for accomplishing the above object consists of the following (1)-(7).

(1) A novel membrane protein polypeptide containing an amino acid sequence shown in sequence No. 1 of the sequence table or a part of the amino acid sequence and being expressed on the synovial membrane of patients with rheumatoid arthritis.

(2) A DNA encoding a polypeptide containing an amino acid sequence shown in sequence No. 1 of the sequence table or a part of the amino acid sequence.

(3) The DNA according to the above (2), characterized by containing a base sequence which hybridizes the base sequence shown in sequence No. 2 of the sequence table or a base sequence derived from said base sequence having at least one amino acid residue substituted, removed or added partially.

(4) A recombinant vector containing the DNA according to the above (2) or (3).

(5) A prokaryotic or eukaryotic host cell, characterized by being transformed with the recombinant vector according to the above (4).

(6) A method for producing the polypeptide containing an amino acid sequence shown in sequence No. 1 of the sequence table or a part of the amino acid sequence, characterized by culturing the host cell according to the above (5).

(7) A monoclonal antibody recognizing a polypeptide containing an amino acid sequence shown in sequence No. 1 of the sequence table or a part of the amino acid sequence.

Subsequently, the present invention will be described in detail.

The monoclonal antibody of the present invention may be prepared in the following manner essentially.

That is, the antibody of the present invention may be prepared by using a synovial cell derived from patients with rheumatoid arthritis (RA) having pre-B cell growth-supporting ability as an antigen, immunizing it according to an ordinary immunization method, cell-fusing the immunized cell according to an ordinary cell fusion method and cloning the fused cell according to an ordinary cloning method.

More specifically, as a preferable method for producing the monoclonal antibody of the present invention may be exemplified a method comprising using the cell line SynSV6-14, derived from the synovial membrane of patients with rheumatoid arthritis (RA) and established as a culture cell, as the above-mentioned antigen, fusing the plasma cell (immunocyte) of a mammal immunized with said antigen with a myeloma cell of a mammal such as a mouse, cloning the obtained fused cell (hybridoma), selecting clones producing the antibody of

the present invention recognizing SynSV6-14 of them, and culturing them to recover the objective antibody.

In the method for producing the above monoclonal antibody, mammals to be immunized with the antigen are not particularly restricted; it is preferable to select one taking compatibility with a myeloma cell to be used for cell fusion into consideration and generally, a mouse, a rat and a hamster are used.

Immunization is performed according to a general method, for example, by administering cultured cells of the cell line SynSV6-14 derived from the synovial membrane of patients with rheumatoid arthritis (RA) into the peritoneal cavity of a mammal according to injection. More specifically, it is preferable to dilute it with or suspend it in PBS or physiological saline to a proper amount and administer it into an animal several times every 4-21 days, together with an ordinary adjuvant if required. In addition, an ordinary carrier (Schlepper) may be employed on the above administration. As an immunocyte, a splenic cell obtained after the final administration of the above cell line is used preferably.

As a myeloma cell of a mammal as the other parent cell to be fused with the above immunocyte may be preferably used known various cell lines including P3 (P3X63Ag8.653) [J. Immunol., 123:1548 (1978)] , p3-U1 [Current Topics in Micro-biology and Immunology, 81:1-7 (1978)] , NS-1 [Eur. J. Immunol., 6:511-519 (1976)] , MPC-11 [Cell, 8:405-415 (1976)] , SP2/0 [Nature, 276: 269-270 (1978)] , FO [J. Immunol. Meth., 35:1-21 (1980)] , S194 [J. Exp. Med., 148:313-323 (1978)] and R210 [Nature, 277:131-133 (1979)] .

The cell fusion of the above immunocyte with a myeloma cell may be performed essentially according to a known method, for example, a method by Milstein et al. [Methods Enzymol., 73:3-46 (1981)] .

More specifically, the above cell fusion may be performed, for example, in an ordinary nutrition medium in the presence of a fusion-accelerating agent. Examples of the fusion-accelerating agent include polyethylene glycol (PEG) and Sendai virus (HVJ), and moreover, auxiliary agents such as dimethyl sulfoxide may be added properly if required in order to enhance the fusing effect. Regarding the ratios of immunocytes and myeloma cells used, the former is preferably used in an amount 1-10 times that of the latter. Examples of a medium used in the above cell fusion include an RPMI-1640 medium and an MEM medium suitable for the proliferation of the above myeloma cell line and other mediums ordinarily used for the culture of this kind of cell, and in addition, supplementary serum such as fetal calf serum (FCS) may be used together.

Cell fusion is performed by mixing prescribed amounts of the above immunocytes and myeloma cells thoroughly in the above medium, adding a PEG solution preheated to about 37 °C, for example, PEG with an average molecular weight of the order of 1,000-6,000, to the medium ordinarily at a concentration of about 30-60 % (W/V) and mixing them. Subsequently, by repeating the operations of adding proper mediums to them successively and centrifuging the reaction mixture, and removing the supernatants can be formed an objective hybridoma.

Said hybridoma is selected by culturing in an ordinary selective medium, for example, an HAT medium (medium containing hypoxanthine, aminopterin and thymidine). The culture in said HAT medium is continued for a time sufficient for cells other than objective hybridomas (non-fused cells) to die out, ordinarily for several days to several weeks. Subsequently, the screening and monocloning of the hybridomas producing the objective antibody are performed according to an

ordinary limiting dilution analysis.

The thus prepared hybridomas producing the monoclonal antibody of the present invention may be subcultured in an ordinary medium and stored in liquid nitrogen for a long time.

In order to collect the monoclonal antibody of the present invention from said hybridomas may be employed a method comprising culturing said hybridomas according to an ordinary method and obtaining it from the supernatants or a method comprising administering a hybridoma into a appropriate mammal to proliferate and obtaining it from its ascites. The former is suitable for obtaining an antibody with a high purity and the latter is suitable for the mass production of the antibody.

Moreover, the antibody obtained according to the above method may be purified to have a high purity employing an ordinary purification means such as a salting out technique, gel filtration and affinity chromatography.

The thus prepared monoclonal antibody of the present invention makes it possible to identify synovial cells of patients with rheumatoid arthritis (RA) expressing a novel membrane protein of an antigen with a high sensitivity and a high precision according to an ordinary immunological means such as radioimmunoassay (RIA), enzyme immunoassay (EIA) and immunofluorescence analysis.

The gene of the present invention is obtained by preparing mRNA from a synovial cell of patients with rheumatoid arthritis (RA) expressing a membrane protein having human pre-B cell growth-supporting ability, and then converting it into a double-stranded cDNA according to a known method. As a cell used for preparing the mRNA can be mentioned, for example, a cell line SynSV6-14 used as an immune source of a hybridoma RS38, but it



is not limited to the cell line and therefore any type of cells expressing the membrane protein having human pre-B cell growth-supporting ability may be used. Incidentally, SynSV6-8 was used in the present invention.

For the preparation of the total RNA for obtaining mRNA can be employed a method for obtaining the total RNA which consists of performing cesium chloride density-gradient centrifugation after a guanidine thiocyanate treatment [Chirgwin et al., Biochemistry, 18:5294 (1979)] , a method which consists of performing a surfactant treatment and a phenol treatment in the presence of the ribonuclease inhibitor of a vanadium complex [Berger & Birkenmeier, Biochemistry, 18:5143 (1979) ] , and other known methods.

The preparation of mRNA from the total RNA can be accomplished by recovering poly(A)<sup>+</sup>RNA from the total RNA according to, for example, affinity column chromatography using an oligo (dT)-bound carrier, for example, cephalose or cellulose, or a batch method. Besides, poly(A)<sup>+</sup>RNA can be further purified according to sucrose density-gradient centrifugation. In addition, there can be mentioned a method for obtaining poly(A)<sup>+</sup>RNA directly without preparing RNA or a convenient method using a commercially available kit.

In order to obtain a double-stranded cDNA from the thus obtained mRNA, for example, a DNA (cDNA) complementary to mRNA is synthesized by using mRNA as a template, and using an oligo (dT) complementary to a poly-A-chain sited at the 3' end as a primer, and then treating it with reverse transcriptase.

The double-stranded cDNA can be also obtained by degrading mRNA according to an alkaline treatment, subjecting the obtained single-stranded cDNA as a

template to a treatment with reverse transcriptase or DNA polymerase (e.g., Klenow fragment), and then treating it with SI nuclease, or treating it directly with RNase and DNA polymerase [Maniatis et al., Molecular Cloning, Cold Spring Harbor Laboratory (1982) and Gubler & Hoffman, Gene, 25:263 (1983) ] . Nowadays, convenient kits have been on the market, and a double-stranded cDNA can be obtained by using them.

The cDNA library can be obtained by inserting the thus obtained cDNA into a proper vector, for example, an EK-type plasmid vector such as pBR322 and pSC101, and a phage vector such as  $\lambda$  gt10, and then transforming Escherichia coli with said vector (e.g., X1776, HB101, DH1, DH5) or the like (refer, for example, to "Molecular Cloning" above).

On the other hand, host cells of other prokaryotes and eukaryotes can be transformed by using a suitable expression vector in which the double-stranded cDNA obtained according to the above-mentioned method is inserted.

The ligation of the double-stranded cDNA to the vector can be performed by adding a proper chemically-synthesized DNA adapter thereto, and subjecting it with a vector DNA cleaved by means of a restriction enzyme in advance to a treatment with T4 phage DNA ligase in the presence of ATP.

The expression vector of the present invention contains a replicative origin, a selective marker, a promoter located in the upstream region of a gene to be expressed, an RNA splice site and a polyadenylated signal.

As a gene expression promoter in a mammal cell may be used virus promoters such as retrovirus, polyoma virus, adenovirus and simian virus (SV) 40, and promoters derived from cells such as human polypeptide chain elongation factor 1 $\alpha$  (HEF-1 $\alpha$  ). For example,

in case of using a promoter of SV40, it can be performed easily according to a method of Mulligan et al. [Nature, 277:108 (1979)] .

As a replicative origin can be used those derived from SV40 polyoma virus, adenovirus and bovine papilloma virus (BPV), and as a selective marker can be used a phosphotransferase APH (3') II or I (neo) gene, a thymidine kinase (TK) gene, an Escherichia coli xanthine-guanine phosphoribosyl transferase (Ecogpt) gene and a dihydrofolate reductase (DHFR) gene.

In order to express the desired gene using a prokaryotic cell as a host cell, the host cell is transformed with a replicon derived from species capable of being fitted for hosts, namely, a plasmid vector containing a replicative origin and a regulation sequence. A vector which has a marker gene capable of imparting the selectivity of a phenotype to transformed cells is preferable. For example, in case of using Escherichia coli as a host cell, it can be transformed using pBR322, a vector originated from the host cell [Boliver et al., Gene, 2:95 (1975)] . The pBR322 contains an ampicillin resistant gene and a tetracycline resistant gene, and therefore transformants can be identified by utilizing either of these resistant properties.

As a promoter needed for the gene expression of a prokaryotic host cell can be mentioned a promoter of a  $\beta$ -lactamase gene [Chang et al., Nature, 275:615 (1978)] , a lactose promoter [Goedde et al., Nature, 281:544 (1979)] , a tryptophan promoter [Goedde et al., Nucleic Acid Res., 8:4057 (1980)] , a tac promoter and the like preferably; however, it is not limited to them.

As a prokaryotic host cell of hosts to be used in the expression system of the present invention can be

mentioned *Escherichia coli*, *Bacillus subtilis*, *Bacillus thermophilus* and the like preferably; however, it is not limited to them.

In addition, as an eukaryotic host cell can be mentioned eukaryotic microorganisms such as *Saccharomyces cerevisiae*, and cells derived from mammals such as a COS cell, a Chinese hamster ovary (CHO) cell, a C127 cell, a 3T3 cell, a Hela cell, a BHK cell, a namalwa cell and a human fetal renal cell (293 cell) preferably; however, it is not limited to them.

Incidentally, the culture of the transformants of the present invention may be performed by selecting culture conditions suitable for host cells appropriately.

The isolation of a cDNA encoding a membrane protein having pre-B cell growth-supporting ability of the present invention can be performed, for example, by using pre-B cell growth-supporting ability as an index or according to a method such as direct expression cloning using an antibody.

The measurement of pre-B cell growth-supporting ability can be performed by using a murine pre-B cell line DW34 [Eur. J. Immunol., 18:1767 (1988)] . That is, a cell expressing the membrane protein having pre-B growth-supporting ability is cultured until it becomes subconfluent on 24-well plates (preferable density being about 50 %) and a proper amount of radiation is irradiated thereupon, DW34 of  $1$  to  $2 \times 10^3$  per well is added thereto, and cultured in the RPMI-1640 medium containing 10 % FCS under the condition of 5 % CO<sub>2</sub> at 37 °C for about 4 to 6 days. The degree of the enhancement of the growth-supporting ability can be found by examining the number of viable cells of DW34 in each well according to trypan blue dye exclusion.

In the present invention, the desired gene could be cloned by repeating the steps, which consist of

selecting a transformant expressing a membrane protein according to flow cytometry by means of an FACScan using a monoclonal antibody RS38 recognizing the novel membrane protein on the synovial cell of patients with rheumatoid arthritis (RA), preparing a transformant again by sorting the plasmid DNA used for the preparation of the transformant, and then screening the transformant according to flow cytometry.

Specifically, a transduced transformant (293T cell) was cultured on well plates and removed from the plates with PBS containing 0.02 % EDTA, and after the cell was washed with an FACS buffer solution composed of PBS containing 2 % FCS and 0.02 %  $\text{NaN}_3$ , it was reacted with RS38 as a primary antibody. Subsequently, after the unreacted primary antibody was removed by washing it with an FACS buffer solution, it was further reacted with a secondary antibody, an FITC-labeled antibody (FITC-labeled anti-mouse goat Ig antibody), dead cells were stained with propidium iodide, and viable cells were analyzed by an FACScan to select transformants responding strongly to RS38.

Further, the complete length of cDNA (pRS38-BOS) encoding a membrane protein polypeptide having novel pre-B cell growth-supporting ability shown in sequence No. 2 of the sequence table could be obtained by repeating the steps, which consist of treating *Escherichia coli* (DH5) containing the cDNA used for the preparation of transformants responding to the antibody with alkali to select a group of plasmids containing the desired gene, subdividing the group of plasmids into some groups of plasmids, transducing them into 293T cells again, and then selecting transformants according to FACScan analysis using the above-mentioned monoclonal antibody RS38.

Incidentally, the *Escherichia coli* DH5 $\alpha$  strain containing pRS38-pUC19 with the cDNA inserted into the

XbaI cleavage sites of a pUC19 vector was deposited at National Institute of Bioscience & Human Technology, Agency of Industrial Science and Technology in Japan, which is an international depositary authority according to Budapest Treaty on the international recognition of the deposit of microorganisms for the purpose of patent procedure, on October 5, 1993, under the name of Escherichia Coli DH5 $\alpha$  (pRS38-pUC19) with accession No. FERM BP-4434.

Generally, the genes of eukaryotes are thought to show polymorphism as known according to human interferon genes [e.g., Nishi et al., J. Biochem., 97: 153 (1985)] , and in some cases at least one amino acid is substituted according to this polymorphism, and in other cases amino acids do not change at all though there are changes in the DNA sequence.

Further, it is probable that some polypeptides having at least one more or less amino acid than the amino acid sequence shown in sequence No. 1 of the sequence table, or some polypeptides substituted with at least one amino acid may also have the same function as that of the novel membrane protein of the present invention (pre-B cell growth-supporting ability). Actually, for example, it has been already known that the polypeptide obtained from a human interleukin-2 (IL-2) gene, in which a DNA sequence corresponding to cysteine is converted to a sequence corresponding to serine, also holds an IL-2 activity [Wang et al., Science, 224:1431 (1984)] .

Moreover, a known protein gene and a gene shown in sequence No. 2 of the sequence table can be ligated by means of a proper restriction enzyme or adapter to yield a polypeptide bound to the known protein. As the known protein gene can be mentioned immunoglobulin, and it may be bound to a Fc portion thereof using the gene shown in sequence No. 2 of the sequence table instead

of the variable region site thereof [ (Zettlmeissl et al., DNA AND CELL BIOLOGY, 9:347-353 (1990) ) ] .

Furthermore, in case of expressing a polypeptide in eukaryotic cells, glycosylation occurs in many cases, and the glycosylation can be regulated according to the conversion of at least one amino acid; in this case, too, it may have the same function as that of the novel membrane protein polypeptide of the present invention. Therefore, even the genes in which the site encoding the membrane protein polypeptide of the present invention are modified artificially according to various methods as above and polypeptides can be included in the present invention so far as the polypeptides obtained from the genes have the same function as that of the membrane protein polypeptide of the present invention.

Moreover, it goes without saying that genes to be hybridized with genes shown in sequence No. 2 of the sequence table and polypeptides are also included in the present invention so far as the polypeptides expressed from the genes have the same function as that of the membrane protein polypeptide of the present invention (pre-B cell growth-supporting ability).. In this case, hybridization may be carried out according to employing ordinary hybridization conditions (for example, refer to the above-mentioned "Molecular Cloning").

The desired homogeneous and purified soluble membrane protein polypeptide having pre-B cell growth-supporting ability can be obtained by culturing a transformant transformed with a gene encoding the polypeptide, solubilizing the yielded polypeptide with a proper detergent, subjecting the resultant polypeptide to separation and purification. Preferable examples of the detergent include Nonidet P-40 (NP-40), Sodium Dodecyl Sulphate (SDS), Triton X-100, Tween 20 and the like.

In addition, soluble membrane proteins can be also

prepared according to gene engineering. Namely, as shown in Fig. 3, since RS38 is guessed to be a cell membrane through-type protein having a cell membrane through domain and an intracellular domain at the side of the N terminal, soluble RS38 with the 49th Asn of sequence No. 1 of the sequence table as the N end can be prepared by employing a PCR-mutagenesis method [M. Kamman et al., Nucl. Acids Res., 15:5404 (1989)]. In this case, as a signal sequence may be used known ones and examples thereof include the signal sequence of Bst-1 (Japanese Patent Application No. 5-141178/1993) and that of G-CSF (Japanese Patent Publication No. 2-5395/1990).

As a means of separation and purification of the membrane protein polypeptide, a method to be used in the case of ordinary protein can be employed; for example, the membrane protein polypeptide of the present invention can be separated and purified properly by selecting and combining various types of chromatography such as affinity chromatography using the above-mentioned monoclonal antibody, ultrafiltration, salting out, dialysis and the like.

#### Brief Description of Drawings

Fig. 1 shows the results (photograph according to agarose gel electrophoresis) of analyzing a gene obtained in Examples of the present invention according to northern blotting analysis.

Fig. 2 shows the growth ability of the mouse pre-B cell line DW34 of a membrane protein obtained in Examples of the present invention.

Fig. 3 shows the results of analyzing the hydrophobic region and the hydrophilic region of a gene obtained in Examples of the present invention by means of a DNA analysis software Gene Works.



The present invention will be described in detail according to Referential Examples and Examples hereinafter, although the present invention is not limited to these Examples.

#### Referential Example 1

##### Establishment of a Stromal Cell Line Derived from Healthy Donors

Stromal cells derived from healthy donors were electroporated with a pAct-SVT plasmid containing an SV40 large T antigen cDNA and a chick  $\beta$ -actin promoter [BBRC, 186:129-134 (1992)] by means of a Gene Pulser (manufactured by BioLad). Namely, 0.8 ml of an aliquot of the stromal cells of  $1 \times 10^7$  cells/ml derived from healthy donors in PBS were mixed with 10  $\mu$ g of the plasmid, and the mixture was incubated on ice for 10 minutes, subjected to electroporation under the conditions of 250 V and at an electrostatic capacity of 250  $\mu$ F, further incubated on ice for 10 minutes, suspended in the RPMI-1640 medium (manufactured by GIBCO) containing 10 % FCS (manufactured by Bioproducts), and cultured in a 10-centimeter culture dish. The culture medium was changed every three days, and colonies of well-grown adhesive cells were harvested about 2 weeks later with a small piece of filter paper impregnated with trypsin to obtain a stromal cell line (NFSV1-1) derived from the bone marrow of healthy donors [J. Immunol., 149:4088 (1992)] .

#### Referential Example 2

##### Establishment of Synovial Cell Lines Derived from Patients with Rheumatoid Arthritis (RA)

Synovial cells derived from patients with rheumatoid arthritis (RA) were electroporated with a pAct-SVT plasmid containing an SV40 large T antigen cDNA and a chick  $\beta$ -actin promoter [BBRC, 186:129-134

(1992)] by means of a Gene Pulser (manufactured by BioLad). Namely, 0.8 ml of an aliquot of the synovial cells of  $1 \times 10^7$  cells/ml derived from patients with RA in PBS were mixed with 10  $\mu$ g of the plasmid, and the mixture was incubated on ice for 10 minutes, subjected to electroporation under the conditions of 250 V and at an electrostatic capacity of 250  $\mu$ F, further incubated on ice for 10 minutes, suspended in the RPMI-1640 medium (manufactured by GIBCO) containing 10 % FCS (manufactured by Bioproducts), and cultured in a 10-centimeter culture dish. The culture medium was changed every three days, and colonies of well-grown adhesive cells were harvested about 2 weeks later with a small piece of filter paper impregnated with trypsin to obtain synovial cell lines (SynSV6-8 and SynSV6-14) derived from patients with rheumatoid arthritis (RA).

### Best Mode for Carrying Out the Invention

### Example 1

## 1) Antigen and Immunization

The cells were treated with 0.02 % EDTA and PBS, and recovered from a culture flask of the incubator

according to pipetting. The cells were suspended into the RPMI medium at a rate of about  $1 \times 10^7$  cells/ml, and immunized to a BALB/C mouse (4-week old, female, manufactured by S. L. C. of Japan). In the initial immunization, about  $1 \times 10^7$ /ml cells were injected into the peritoneal cavity of the mouse, and 2 to 3 weeks later,  $1 \times 10^7$ /ml cells were injected as additional immunization. Further, at intervals of 2 to 3 weeks,  $1 \times 10^7$ /ml cells were injected 2 to 3 times as additional immunization, and 3 days after the final immunization, the mouse was sacrificed and the spleen was obtained for fusion.

## 2) Cell fusion

After the spleen extirpated from one mouse was cut to pieces, isolated spleen cells were centrifuged, suspended into RPMI-1640 medium (manufactured by GIBCO), and washed sufficiently. On the other hand,  $1 \times 10^7$  cells obtained by culturing a mouse-myeloma cell line P3X63Ag8.653 [J. Immunol., 123:1548 (1979)] in the DMEM medium (manufactured by GIBCO) containing 10 % fetal calf serum (FCS, manufactured by FILTRON) were washed in the above DMEM medium similarly, and were introduced into a centrifugal tube together with  $1 \times 10^8$  of said spleen cells and mixed, and then were subjected to cell-fusion with polyethylene glycol 1500 (manufactured by Boehringer) according to ordinary procedure [Clin. Exp. Immunol., 42:458-462 (1980)] .

The obtained fused cells were introduced into 96-well plates in the DMEM medium containing 10 % FCS, and cultured in an incubator containing 5 %  $\text{CO}_2$ , at 37 °C.

From the following day, the medium was replaced with the HAT selective medium (complete RPMI-1640 medium containing  $1.0 \times 10^{-4}$  M hypoxanthine,  $4.0 \times 10^{-7}$  M aminopterin and  $1.6 \times 10^{-5}$  M thymidine having 10 % FCS and 50  $\mu$  M 2-mercaptoethanol added thereto) slowly, and the culture was continued. After the culture was

initiated, half of the supernatant was replaced with a new HAT medium 2 times per week, and the culture was continued to maintain proliferation.

The thus obtained fused cells were cloned according to limiting dilution analysis.

That is, using the antibodies in the culture supernatant obtained by culturing the above fused cells, responsency with the antigen was examined, and clones having strong responsency with the antigen alone were obtained according to ordinary procedure employing limiting dilution analysis.

So as to perform the formation of clones, the above hybridoma and the spleen cells of a BALB/C mouse were prepared in prescribed amounts, inoculated onto 96-well plates at a rate of 1 to 10 hybridoma(s) per well, and cultured in an incubator containing 5 % CO<sub>2</sub>, at 37 °C. The operation of cloning hybridomas grown was repeated in the same manner according to ordinary limiting dilution analysis until they became a single clone theoretically. Clones yielding the desired antibody were screened using the above antigen.

### 3) Screening

The screening of fused cells (hybridomas) was performed according to indirect fluorescent antibody technique, flow cytometry analysis by means of a flow cytometer.

The screening of clones yielding the objective antibody was performed using a) SynSV6-14 (antigen for immunization) and b) the bone marrow stromal cell line NFSV1-1 derived from healthy donors as target cells. Namely, after immunizing the synovial cell line (SynSV6-14) derived from patients with rheumatoid arthritis (RA) having pre-B cell growth-supporting ability to a BALB/C mouse, the screening of monoclonal antibodies responding to SynSV6-14 but not responding to the bone marrow stromal cell line (NFSV1-1) derived from healthy

donors having no pre-B cell growth-supporting ability was performed as follows. The first screening was performed using SynSV6-14, a cell to be subjected to reaction, as an antigen for immunization. First of all, culture supernatants responding to SynSV6-14 were selected with a view to selecting fused cell clones responding to said SynSV6-14, and then a primary screening was performed.

Namely, cells suspended in a reaction buffer (PBS containing 2 % FCS and 0.02 % NaN<sub>3</sub>) were suspended into 20  $\mu$ l of a hybridoma culture supernatant (about 5 x 10<sup>5</sup>/20  $\mu$ l), and reacted at 4 °C for 20 minutes.

They were washed with the above buffer twice, and the FITC-labeled anti-mouse Ig goat antibody (manufactured by Cappel) was added thereto, and the mixture was incubated for 20 minutes. After the reaction product was washed three times, it was analyzed by means of a flow cytometer (FACScan, manufactured by Becton Dickinson).

Subsequently, the bone marrow stromal cell line NFSV1-1 derived from healthy donors was used as a cell to be subjected to reaction and analyzed by means of a flow cytometer as above. A hybridoma yielding antibodies responding more strongly to SynSV6-14 was obtained according to it.

Thus, a hybridoma (RS38) yielding antibodies responding to SynSV6-14 but not responding to the bone marrow stromal cell line NFSV1-1 derived from healthy donors was isolated. The antibodies yielded by the hybridoma were IgM,  $\kappa$ -type.

Incidentally, the hybridoma yielding the above monoclonal antibody RS38 is a novel fused cell prepared from a BALB/C mouse spleen cell and mouse-myeloma P3X63Ag8.653 as parent cells and was deposited at National Institute of Bioscience & Human Technology, Agency of Industrial Science and Technology in Japan,

which is an international depositary authority according to Budapest Treaty on the international recognition of the deposit of microorganisms for the purpose of patent procedure, on October 5, 1993, under the name of Mouse-Mouse hybridoma RS38 with accession No. of FERM BP-4433.

#### 4) Purification of Antibodies

The fused cells prepared in the above 2) were cultured according to ordinary procedure, and the antibodies yielded in the culture supernatant were purified according to ordinary procedure.

That is, hybridomas were collected from the wells with the highest antibody titer to the above antigen, and one well in which the growth of cells could be recognized was taken out, and the obtained culture cells were expanded into a tissue culture flask under the conditions of 5 % CO<sub>2</sub> at 37 °C and were grown. The obtained cells were injected into the peritoneal cavity of a BALB/C mouse (6-week old, female, manufactured by S. L. C. of Japan) with pristane dosed. And 10 to 14 days after, the ascites was collected, salted out with 50 % ammonium sulfate, dialyzed with PBS and purified with a QAE column. The antibodies were further salted out and dialyzed sufficiently to obtain a purified product of about 6 mg/ml.

#### Example 2

##### Properties of the monoclonal antibodies

#### 1) Distribution of Cell Lines

The distribution of the RS38 antigen in various cell lines (various cell lines shown in Table 1) was analyzed by means of an FACScan. That is, each of various cell lines was suspended in an FACS buffer solution comprising PBS containing 2 % FCS and 0.02 % NaN<sub>3</sub>, in the presence of 10 µg/ml of the mouse monoclonal antibody RS38 obtained as a primary antibody

in Example 1, and incubated on ice for 20 minutes. After washed with an FACS buffer solution twice, the resultant product was further incubated on ice for 15 minutes using an FITC-labeled anti-mouse Ig goat antibody (manufactured by Cappel) as a secondary antibody. Propidium iodide (PI) was added therein so that the final concentration of it became  $1\mu$  g/ml, and the mixture was further incubated on ice for 5 minutes.

After the resultant product was washed with the FACS buffer solution three times, the cells were analyzed with light scattering measurement and viable cells alone were subjected to analysis by means of an FACScan (manufactured by Becton Dickinson). As controls were used mouse monoclonal antibodies SG2 and RF3 (Japanese Patent Application No. 5-141178/1993). The results are shown in Table 1. As is apparent from Table 1, it was revealed that the distribution of the RS38 is different from that of SG2 and RF3.

Table 1

Results of FACS analysis on Cell Lines

Origen	Cell Line	SG2/RF3	RS38
T cell line	MOLT-4	—	+ / —
	JURKAT	+	—
B cell line	Raji	—	—
	Paudi	—	—
	CL4	—	++
	Ramos	—	—
Myeloid	U937	+	+
	K562	—	+
	HL60	—	+ / —
	MO7	—	+
Miscellaneous	HepG2	—	+ / —
	T24	—	+ / —
	SKWG-4 (P122)	—	+ / —
	Hela	—	+ / —



### Example 3

#### 1. Preparation of a cDNA Library

##### 1) Preparation of Poly (A)<sup>+</sup>RNA

The preparation of poly (A)<sup>+</sup>RNA from the synovial cell line SynSV6-8 derived from patients with rheumatoid arthritis (RA) was performed using a Fast Track<sup>TM</sup> mRNA isolation kit version 3.2 (manufactured by Invitrogen).

That is, SynSV6-8 cells for twenty 10-centimeter culture dishes were homogenized, and then the total RNA was prepared according to the procedure attached to the kit. Further, the poly (A)<sup>+</sup>RNA was purified by means of oligo d(T) cellulose attached to the kit according to the procedure attached to the kit.

##### 2) Construction of the cDNA Library

A double-stranded cDNA was synthesized using the above poly (A)<sup>+</sup>RNA of 5  $\mu$ g as a material according to the procedure attached to a cDNA synthesis kit, Time Saver<sup>TM</sup> cDNA synthesis kit (manufactured by Pharmacia), and a BstXI adapter (manufactured by Invitrogen) was ligated thereto by means of a DNA ligation kit (manufactured by Takara Shuzo) according to the procedure attached to the kit. The removal of the free BstXI adapter was performed by means of the Size Sep 400 Spin Column attached to the kit according to the procedure attached to the kit to obtain about 100  $\mu$ l of an adapter-ligated double-stranded cDNA.

And then, of about 100  $\mu$ l of the thus prepared adapter-ligated double-stranded cDNA, 2  $\mu$ l were used in one ligation reaction, and a cDNA library was constructed by ligating it with a pEF-BOS vector [Nuc. Acid Res., 18:5322 (1990)] treated in advance with a restriction enzyme BstXI and alkali phosphatase (manufactured by Takara Shuzo) by means of a cDNA ligation kit (manufactured by Takara Shuzo). The constructed cDNA library was transformed into the Escherichia coli cell line DH5 (manufactured by Toyobo)

and was presumed to be an independent clone with the total size of about  $2 \times 10^5$ . Fifty pools, each pool comprising 2,000 to 4,000 clones of transduced *Escherichia coli*, were prepared and then used in the following tests.

## 2. Cloning according to Direct Expression Method

### 1) Transfection into 293T Cells

The amplification of a cDNA was performed by culturing the above pooled *Escherichia coli* in the LB medium containing 50  $\mu$ g/ml of ampicillin [Molecular Cloning: A Laboratory Manual, Sambrook et al., Cold Spring Harbor Laboratory Press (1989)], and a plasmid DNA was recovered from the *Escherichia coli* according to an alkaline method [Molecular Cloning: A Laboratory Manual, Sambrook et al., Cold Spring Harbor Laboratory Press (1989)]. The degree of the purification of the obtained plasmid DNA was enhanced by repeating ultracentrifugation according to cesium chloride/ethidium bromide density-gradient centrifugation, and the purified plasmid DNA was transfected to a 293T cell [cell line prepared by transfecting an SV40 large T antigen cDNA into a 293 cell (Transformed primary embryonal kidney, human ATCC CRL 1573)] according to a calcium phosphate method.

Namely, 2  $\mu$ g of the purified plasmid DNA was dissolved into 100  $\mu$ l of a buffer solution containing 1 mM of Tris-HCl and 0.1 mM of EDTA, and after 14  $\mu$ l of 2M CaCl<sub>2</sub> were added thereto, the resultant mixture was mixed with a buffer solution composed of 50 mM of HEPES (pH: 7.1), 280 mM of NaCl and 1.5 mM of sodium phosphate slowly, and then the obtained mixture was incubated at room temperature for 30 minutes and added to the 293T cells in 24-well plates. The 293T cells were cultured in the DMEM (manufactured by GIBCO) medium containing 10 % fetal calf serum (FCS, manufactured by Bioproducts) under the conditions of 37 °C and 5 % CO<sub>2</sub>.

for 2 days.

## 2) Analysis according to FACScan

The transduced 293T cells were removed from the 24-well plates in PBS containing 0.02 % EDTA, and washed with an FACS buffer solution comprising PBS containing 2 % FCS and 0.02 % NaN<sub>3</sub>, twice, and then suspended in 20  $\mu$ l of the FACS buffer solution in the presence of 10  $\mu$ g/ml of the above-mentioned monoclonal antibody RS38 as a primary antibody and incubated on ice for 20 minutes.

After they were washed with the FACS buffer solution twice, they were further incubated on ice for 15 minutes, using an FITC-labeled anti-mouse Ig goat antibody (manufactured by Cappel) as a secondary antibody. Propidium iodide (PI) was added therein so that the final concentration of it became 1  $\mu$ g/ml, and the mixture was further incubated on ice for 5 minutes, washed with the FACS buffer solution three times, and the cells were analyzed with light scattering measurement and viable cells alone were subjected to analysis by means of an FACScan (manufactured by Becton Dickinson).

## 3) Cloning of the cDNA Library

The plasmid DNAs recovered from Escherichia coli of 2,000 to 4,000 clones as one pool according to an alkaline method were transfected to 293T cells according to the above method, and the transfected cells were subjected to screening according to the above FACS analysis. A peak strongly stained with the mouse monoclonal antibody RS38 was recognized in the 25th pool of the 293T cells. The plasmid DNA was transduced into Escherichia coli DH5 (manufactured by GIBCO BRL) again, and it was inoculated onto an LB agar plate containing 50  $\mu$ g/ml of ampicillin.

About 2,000 clones forming colonies were inoculated one by one onto an agar plate with its bottom divided into a net-like state so that the position of a clone

inoculated was recognized at a rate of 100 clones per plate, and two series of each plate were prepared. 20 pools, each pool comprising 100 clones, were prepared in the same way, and *Escherichia coli* was cultured in the LB medium containing 50  $\mu$ g/ml of ampicillin. After plasmid DNAs were recovered according to an alkaline method, they were transfected to 293T cells according to a calcium phosphate method, and the transfected cells were subjected to FACScan analysis in the same manner as above. As a result of the FACScan analysis, 100 clones of *Escherichia coli* were isolated one by one from one pool recognized to be positive, and each clone was cultured, and then plasmid DNAs were recovered according to an alkaline method. Each plasmid DNA was transfected to 293T cells according to a calcium phosphate method, and FACScan analysis was performed in the same manner as above to obtain a single positive clone, which was designated as pRS38-BOS.

The clone was subjected to a sequence reaction using an Auto Read sequencing kit (manufactured by Pharmacia) and an Auto Cycle sequencing kit (manufactured by Pharmacia) according to the procedure attached to the kits, and the determination of its DNA sequence was performed by means of an A. L. F. <sup>TM</sup> DNA sequenator (manufactured by Pharmacia). As a result, it was a gene with the full length of 996 bp (sequence No. 2 of the sequence table) to be presumed to encode a sequence of the 180 amino acid residues from the longest open reading frame, and as a result of performing homology retrieval according to data bases SWISS PLOT and NBRL using a gene analysis software Genetex, it was recognized to be a novel gene.

#### 4) Investigation of Expression according to Northern Blotting Analysis

Preparation of poly (A)<sup>+</sup>RNA from the synovial cell line SynSV6-14 derived from patients with RA, the bone

marrow stromal cell line RASV5-5 derived from patients with RA and the bone marrow stromal cell line NFSV1-1 derived from healthy donors was performed by using a Fast Track™ mRNA isolation kit version 3.2 (manufactured by Invitrogen) to perform northern blotting analysis (Molecular Cloning; A Laboratory Manual, Sambrook et al., Cold Spring Harbor Laboratory Press, 1989). That is, cell lines cultured in ten 10-centimeter culture dishes were recovered and homogenized, and then the total RNA was prepared according to the procedure attached to the kit. Further, the poly (A)+RNA was purified by means of oligo d(T) cellulose attached to the kit according to the procedure attached to the kit.

Labelling a probe was performed by using a Multiprime DNA labelling system (manufactured by Amersham). That is, a 315bp fragment was prepared by digesting an insert believed to encode RS38 inserted into the BstXI site of pEF-BOS with a restriction enzyme HindIII and then a labelled probe was prepared according to the procedure attached to the kit. The poly (A)+RNA prepared from SynSV6-14, RASV5-5 and NFSV1-1, 3  $\mu$ g per lane, was subjected to agarose gel electrophoresis, and then blotting was performed according to a capillary method by using a Gene Screen Plus™ (manufactured by Du Pont) as a hybridization transfer membrane. Hybridization was performed at 65 °C overnight by using a Gilbert & Church buffer comprising 0.5 M NaPO<sub>4</sub>, 1mM EDTA, 7 % SDS and 1 % BSA, pH 7.0. After the completion of hybridization, the membrane was washed with 2 x SSC at room temperature four times and further with 0.1 x SSC and 0.1 % SDS at 55 °C twice, and then placed face to face with an X-ray film to perform autoradiography overnight. As a result, an apparent band of about 1.0 kb was detected on SynSV6-14 as shown in Fig. 1.

### 3. Expression by a BALB3T3 Cell

The novel molecule was transfected into a BALB3T3 cell and the expression in mammalian cells was examined.

Namely, 20  $\mu$ g of pRS38-BOS and 2  $\mu$ g of pSV2neo which has a neomycin-resistant gene [P. J. Souethem and P. Berg, J. Mol. Appl. Genet., 1:327 (1982)] were added into 0.8 ml of aliquot of  $1 \times 10^7$  cells/ml, and incubated on ice for 10 minutes, and then subjected to transfection by means of a Gene Pulser (manufactured by BioLad) under the conditions of 250 V and an electrostatic volume of 250  $\mu$ F to accomplish transduction simultaneously.

Further, after it was incubated on ice for 10 minutes, the cell was suspended in the DMEM medium (manufactured by GIBCO) containing 2 mg/ml of G418 and 10 % FCS (manufactured by Bioproducts) and cultured in 24-well plates. The replacement of a culture medium was performed every three days, and about 2 weeks later, transformed cell lines BALB3T3 38-1, 38-3 and 38-9 responding to the above mouse monoclonal antibody RS38 were obtained from the well forming a single colony of well-grown adhesion cells having neomycin resistance. In addition, as a control cell was obtained a transformed cell line BALB3T3 38-4 not responding to RS38 but having neomycin resistance.

### 4. Biological Properties of the Novel Molecule

#### 1) Growth-Supporting of Mouse Pre-B Cell Lines

The biological properties of the novel molecule were analyzed using a mouse pre-B cell line DW34 growing dependent on stromal cells with the number of grown cells as an index according to the following method.

First of all, transduced cell lines BALB3T3 38-1, 38-3 and 38-9 and control cell lines BALB3T3 and BALB3T3 38-4 were cultured in 24-well plates until they

became subconfluent, radiation of 30 Gy was irradiated thereupon, DW34 of  $2 \times 10^3$  per well was added therein, and the resultant product was cultured in the RPMI-1640 (manufactured by GIBCO) medium containing 10 % FCS (manufactured by Bioproducts) under the conditions of 37 °C and 5 % CO<sub>2</sub> for 4 days. The number of viable cells of DW34 in each well was counted with trypan blue dye exclusion to analyze a growth-supporting ability. As a result, the growth of DW34 was enhanced in the transduced cell lines BALB3T3 38-1, 38-3 and 38-9 in comparison with control cell lines BALB3T3 and BALB3T3 38-4 as shown in Fig. 2.

## 5. Physicochemical Properties of the Novel Molecule

### 1) Analysis of the N Terminal

The analysis of the hydrophobic region and the hydrophilic region of the gene obtained as above was performed by using a DNA analysis software Gene Works.

The results of the analysis are shown in Fig. 3. As a result, the region of 28 amino acid residues from the 21st Lys to the 48th Ala shown in sequence No. 1 of the sequence table had the highest hydrophobic properties, and therefore it was presumed to be type of a protein to pass through the cell membrane, having a domain passing through cell membrane and an intracellular domain at the side of the N terminal of mature protein.

### Industrial Applicability

As described in detail above, the present invention relates to a gene encoding a novel membrane protein polypeptide having pre-B cell growth-supporting ability, a vector containing said gene, transformants transformed by said vector, and a method for producing a novel membrane protein having pre-B cell growth-supporting ability using said gene, and therefore the gene of the present invention is capable of encoding the membrane protein on the synovial cell derived from

patients with rheumatoid arthritis (RA).

The homogeneous and purified novel membrane protein polypeptide can be produced in large quantities by inserting the gene of the present invention into a suitable vector and then transforming ordinary host cells, and in addition, monoclonal antibodies recognizing said membrane protein polypeptide can be produced by using the synovial cell derived from patients with rheumatoid arthritis (RA) as an antigen for immunization. Thus, according to the present invention, it becomes possible to identify rheumatoid arthritis (RA) and also prepare reagents for the clinical diagnosis thereof.



【 SEQUENCE TABLE】

SEQ ID NO: 1

SEQUENCE LENGTH: 180

SEQUENCE TYPE: Amino acid

TOPOLOGY: Linear

MOLECULE TYPE: Peptide

SEQUENCE DESCRIPTION:

Met	Ala	Ser	Thr	Ser	Tyr	Asp	Tyr	Cys	Arg	Val	Pro	Met	Glu	Asp	Gly
				5					10					15	
Asp	Lys	Arg	Cys	Lys	Leu	Leu	Leu	Gly	Ile	Gly	Ile	Leu	Val	Leu	Leu
			20					25					30		
Ile	Ile	Val	Ile	Leu	Gly	Val	Pro	Leu	Ile	Ile	Phe	Thr	Ile	Lys	Ala
		35					40				45				
Asn	Ser	Glu	Ala	Cys	Arg	Asp	Gly	Leu	Arg	Ala	Val	Met	Glu	Cys	Arg
		50				55					60				
Asn	Val	Thr	His	Leu	Leu	Gln	Gln	Glu	Leu	Thr	Glu	Ala	Gln	Lys	Gly
	65			70				75					80		
Phe	Gln	Asp	Val	Glu	Ala	Gln	Ala	Ala	Thr	Cys	Asn	His	Thr	Val	Met
			85					90					95		
Ala	Leu	Met	Ala	Ser	Leu	Asp	Ala	Glu	Lys	Ala	Gln	Gly	Gln	Lys	Lys
		100					105					110			
Val	Glu	Glu	Leu	Glu	Gly	Glu	Ile	Thr	Thr	Leu	Asn	His	Lys	Leu	Gln
		115					120					125			
Asp	Ala	Ser	Ala	Glu	Val	Glu	Arg	Leu	Arg	Arg	Glu	Asn	Gln	Val	Leu
	130						135				140				
Ser	Val	Arg	Ile	Ala	Asp	Lys	Lys	Tyr	Tyr	Pro	Ser	Ser	Gln	Asp	Ser
	145					150				155				160	
Ser	Ser	Ala	Ala	Ala	Pro	Gln	Leu	Leu	Ile	Val	Leu	Leu	Gly	Leu	Ser
			165					170					175		
Ala	Leu	Leu	Gln												

SEQ ID NO: 2  
 SEQUENCE LENGTH: 996  
 SEQUENCE TYPE: Nucleic acid  
 STRANDEDNESS: Double  
 TOPOLOGY: Linear  
 MOLECULE TYPE: cDNA to mRNA  
 METHOD FOR DETERMINING FEATURES: E  
 SEQUENCE DESCRIPTION:

GTGGAATTC ATG GCA TCT ACT TCG TAT GAC TAT TGC AGA GTG CCC ATG GAA	51
GAC GGG GAT AAG CGC TGT AAG CTT CTG CTG GGG ATA GGA ATT CTG GTG	99
CTC CTG ATC ATC GTG ATT CTG GGG GTG CCC TTG ATT ATC TTC ACC ATC	147
AAG GCC AAC AGC GAG GCC TGC CGG GAC GGC CTT CGG GCA GTG ATG GAG	195
TGT CGC AAT GTC ACC CAT CTC CTG CAA CAA GAG CTG ACC GAG GCC CAG	243
AAG GGC TTT CAG GAT GTG GAG GCC CAG GCC GCC ACC TGC AAC CAC ACT	291
GTG ATG GCC CTA ATG GCT TCC CTG GAT GCA GAG AAG GCC CAA GGA CAA	339
AAG AAA GTG GAG GAG CTT GAG GGA GAG ATC ACT ACA TTA AAC CAT AAG	387
CTT CAG GAC GCG TCT GCA GAG GTG GAG CGA CTG AGA AGA GAA AAC CAG	435
GTC TTA AGC GTG AGA ATC GCG GAC AAG AAG TAC TAC CCC AGC TCC CAG	483
GAC TCC AGC TCC GCT GCG GCG CCC CAG CTG CTG ATT GTG CTG CTG GGC	531
CTC AGC GCT CTG CTG CAG TGAGATCCCA GGAAGCTGGC ACATCTTGGA AGGTCCGTCC	589
TGCTCGGCTT TTCGCTTGAA CATTCCCTTG ATCTCATCAG TTCTGAGCGG GTCATGGGGC	649
AACACGGTTA GCGGGGAGAG CACGGGGTAG CCGGAGAAGG GCCTCTGGAG CAGGTCTGGA	709
GGGGCCATGG GGCAGTCCTG GGTGTGGGGA CACAGTCGGG TTGACCCAGG GCTGTCTCCC	769
TCCAGAGCCT CCCTCCGGAC AATGAGTCCC CCCTCTTGTC TCCCACCCTG AGATTGGGCA	829
TGGGGTGCGG TGTGGGGGGC ATGTGCTGCC TGTGTATTATG GGTTTTTTTT GCGGGGGGGG	889
TTGCTTTTTT CTGGGGTCTT TGAGCTCCAA AAAATAAACA CTTCTTTTGA GGGAGAGCAA	949
AAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAGAATTC CACCACA	996